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Perfluorodecanoic acid stimulates NLRP3 inflammasome assembly in gastric cells

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Perfluorodecanoic acid (PFDA), a perfluorinated carboxylic acid, presents in the environment and accumulates in human blood and organs, but its association with tumor promotion are not clear. Given that inflammation plays a significant role in the development of gastric malignancies, we evaluated the effects of PFDA on activation of the inflammasome and inflammation regulation in the gastric cell line AGS. When added to cell cultures, PFDA significantly stimulated IL-1 β and IL-18 secretion and their mRNA levels compared with control cells. By RT-PCR and western-blot we found that up-regulation of NLRP3 were associated with promotion of IL-1 β and IL-18 production. Then expression variation of cIAP1/2, c-Rel and p52 were analyzed, the results demonstrated raised mRNA expression in all the tested genes concomitant with enhanced inflammasome activity after exposure to PFDA. Assays with cIAP2 siRNA and NF κ B reporter provided additional evidence that these genes were involved in PFDA-induced inflammasome assembly. Furthermore, increased secretion of IL-1 β and IL-18 were detected in stomach of PFDA-treated mice, disorganized alignment of epithelial cells and inflammatory cell infiltration were also observed in the stomach tissues upon PFDA treatment. This study reports for the first time that PFDA regulates inflammasome assembly in human cells and mice tissues.

Gastric cancer is the second leading cause of cancer death worldwide and accounts for about 10% of all invasive cancers¹. Although the pathogenesis of gastric cancer is not completely understood, it is worth noting that a positive correlation of gastric cancer with environmental pollution has been confirmed^{2–4}. As an illustration, the total number of cases and deaths from gastric cancer have increased concomitant with extensive demographic changes and ongoing increase of environmental pollution in China.

Perfluorodecanoic acid (PFDA) is a highly toxic perfluorinated fatty acid (PFCA)⁵, a kind of persistent environmental pollutant. PFDA is present in air, food, and water, especially in China, where 139ng/L PFDA was detected in snow fall in the area around Beijing⁶. PFDA intake is primarily from foods and drinking water^{7,8}, and it can accumulate in human blood and organs. Moreover, its serum elimination half-life lasts several years. PFDA has been reported to induce hypophagia and severe weight loss, bradycardia, hypothermia, and decreased serum thyroid hormone levels in rats^{9,10}. So far, there is certainly a need for more data in the field of PFDA-induced cancers.

While the pathogenesis of gastric cancer is not completely understood, epidemiological studies suggest that chronic inflammation plays a significant role in the development of gastric malignancies¹¹. Emerging evidence indicates that the inflammasome is an essential component of the innate immune system, playing a central role in regulating inflammation¹². The inflammasome complex is composed of a nucleotide binding domain, leucine-rich repeat-containing receptor (NLR), caspase-1 and the ASC (apoptosis-associated speck-like protein containing a caspase recruitment domain) adaptor protein, which is necessary to join NLR and caspase-1¹³. The activation of the inflammasome complex triggers the recruitment and activation of caspase-1, which is necessary for the maturation and release of two important proinflammatory cytokines, IL-1 β and IL-18¹³. Different inflammasomes

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have been described, i.e. NLRP1, NLRP3 and NLRC4. Inflammasomes assembly generally requires NF κ B activation and danger signals, generating active caspase-1 that cleaves the precursor forms of IL-1 β and IL-18 into the mature and active forms. Some evidence suggested that NLRs are closely correlated to cancer occurrence, the level of IL-1 β and IL-18 were found to be significantly elevated in various types of malignancies^{12,14,15}. Therefore, elucidating the mechanisms that involve activation of the inflammasomes and their modulatory effects has become a novel strategy for inflammation regulation and cancer prevention research.

In this study, we found that PFDA induced IL-1 β and IL-18 secretion in culture solution of cell line AGS and mice stomachs. Our results demonstrated that the IL-1 β and IL-18 inductions were associated with activation of NLRP3 inflammasome. This study reports for the first time that PFDA activates inflammasome and promotes gastric inflammation in gastric cells and mice tissues.

Material & methods

Cell line and animal treatment. Human gastric adenocarcinoma cell line AGS, human hepatocellular carcinoma cell lines BEL7402 and human leukemia macrophage cell line THP-1 were all from American type culture collection (ATCC) and maintained in our laboratory. They were cultured in Ham's F-12 medium (HyClone, Utah, USA) containing 10% FCS and 1% penicillin-streptomycin. PFDA was purchased from Sigma Chemical Company (CAS number 335-76-2, St. Louis, MO, USA). Different volumes of PFDA were added into cell culture media for treatments. Six-week-old female Balb/c mice were acclimated and caged in groups of 5 or less. NLRP3 knockout mice were kindly offered by Professor Rongbin Zhou (University of Science and Technology of China). Mice were administered with PFDA at a dose of 25 mg/kg/d in drinking water, 0.014 M dimethyl sulfoxide (DMSO) was utilized as the control. Mice treatment and dissection were performed according to the protocol approved by the Ethics Committee on Animal Experiments of Medical School of Shandong University. Stomachs were harvested 18d after PFDA dosing. Some stomachs were cut into small pieces and mixed with Phosphate-buffered saline in a certain percentage for tissue grinding. The obtained tissue homogenates were used for subsequent Enzyme-linked immunosorbent assay. Some stomachs were Formalin-fixed and paraffin-embedded for Hematoxylin and eosin staining. The rest of stomachs were snap-frozen in liquid nitrogen and stored at -80°C for RNA extraction and quantitative RT-PCR. FuGENEs HD Transfection Reagent (Roche Applied Science, Basel, Switzerland) was used for transfection. All transfections were performed according to the manufacturer's instructions.

RNA extraction and quantitative real-time PCR. Total cellular RNA was extracted with Trizol (Life Technologies, California, USA) according to the protocol provided by the manufacturer. First-strand cDNA was synthesized from 1 μg total cellular or tissue RNA using the RevertAid TM First Strand cDNA Synthesis Kit (Thermo Fisher Scientific, Massachusetts, USA) with random primers. Then cDNA was amplified for quantitative real-time PCR, the specific primers used were as follows: for human cIAP1 (Birc2), forward primer 5'-AATGGAAGATAGCACGAT-3' and reverse primer 5'-CCTTTCTGAGACAGGCAC-3'; cIAP2 (Birc3), forward primer 5'-GCCTGATGCTGGATAACT-3' and reverse primer 5'-GAATAAGAGCCACGGAAA-3'; for human NLRP3, forward primer 5'-TGAACAGCCACCTCACTT-3' and reverse primer 5'-CAACCACAATCTCCGAAT-3'; for human NLRP1, forward primer 5'-CCAGTTTGTGCGAATCCA-3' and reverse primer 5'-CCAACGTAGAACTCCGAGAA-3'; for human NLRC4, forward primer 5'-CAATAGCCGAGCCCTTAT-3' and reverse primer 5'-AGCCAAATCGTCCAAGTC-3'; for human IL-1 β , forward primer 5'-ACAGTGGCAATGAGGATG-3' and reverse primer 5'-TGTAGTG GTGGTCGGAGA-3'; human IL-18, forward primer 5'-ATAGCCAGCCTAGAGGTA-3' and reverse primer 5'-ATCA GGAGGATTCATTTTC-3'; for β -actin, forward primer 5'-AGTTGCGTTACACCCTTTCTTG-3' and reverse primer 5'-CACCTCACCGTTCCAGTTTTT-3'. The real-time PCR reactions were performed at: 95 $^{\circ}\text{C}$, 10sec (denaturation); 55 $^{\circ}\text{C}$, 30sec (annealing); 72 $^{\circ}\text{C}$, 30sec (extension) for 35 cycles. The real-time PCR reactions were performed on the ABI7000 Fast Real-Time PCR System with SYBR Premix Ex Taq TM according to the procedures.

Enzyme-linked immunosorbent assay (ELISA). IL-1 β and IL-18 in culture solution or mouse gastric tissues were detected with human/mouse IL-1 β and IL-18 ELISA Kit (4 A Biotech Co., Ltd, Beijing, China) according to the manufacturer's instructions. Mice treatment and dissection were performed according to the protocol approved by the Ethics Committee on Animal Experiments of Medical School of Shandong University. Briefly, add 100 μl diluted Standard in triplicate or 100 μl sample to each well, then add 50 μl biotin labeled Antibody Working Solution per well. After aspiration and wash for 4 times, incubate for 2 hours at room temperature. Next, add 100 μl Enzyme Binding Working Solution to each well, aspirate and wash 4 times. And then add 100 μl Substrate Solution to each well. Protect from light, incubate for 10–20 minutes at RT. Finally, add 100 μl Stop Solution to each well and read at 450 nm within 30 minutes.

Western Blot Analysis. Western Blot Analysis was performed as described previously¹⁶. Briefly, cell lysates (20 μg /lane) were separated on 10% SDS polyacrylamide gel and then were transferred to a poly (vinylidene fluoride) membrane. NLRP3 (Cat No.TA336883), cIAP1 (sc-271419) and 2 (Cat No.TA800041), c-Rel (Cat No.TA324346) and p52 (sc-7386) protein was detected by a mouse monoclonal IgG (OriGene Co. Ltd, Beijing, China and Santa Cruz Biotechnology Inc., USA) and visualized by the enhanced chemiluminescence system (Amersham, Arlington Heights, IL). The density of the bands was quantitated using the NIH image software package. The intensity of gene expression was judged by the ratio of their expression in PFDA treated groups to their corresponding expression in DMSO groups, and a ratio of more than 1.0 was considered to be an indication of over-expression.

siRNA interference. Chemical modified Stealth siRNA targeting cIAP2 and control siRNA were from RiboBio Co., Ltd. (Guangzhou, Guangdong, China). The sequence for cIAP2 siRNA was 5'-GGAGTTCATCCGTCAGTT-3'. Cells were transfected with siRNA by the Lipofectamine 2000 method (Life Technologies, California, USA).

Hematoxylin and eosin (H&E) staining. Hematoxylin (Code No. ZLI9606), eosin (Code No. ZLI9612), and diaminobenzidine (DAB, Code No. ZLI9632) staining kits were purchased from Zhongshan Golden Bridge Biotechnology (Beijing, China). Mice treatment and dissection were performed according to the protocol approved by the Ethics Committee on Animal Experiments of Medical School of Shandong University. Formalin-fixed, paraffin-embedded mice stomach tissues were cut into 4- μ m thick tissue sections and then stained with hematoxylin and eosin (HE) according to the manufacturer's instructions. The staining images were acquired using a light microscope to observe the pathological alterations of the stomach tissues.

Statistical Data analysis. Data were expressed as mean \pm standard deviation (SD). Differences between three groups were compared using the Student's t-tests and ANOVA. All experiments were repeated at least three times and $P < 0.05$ (*) was considered statistically significant.

Results

PFDA exposure increases the secretion and mRNA levels of IL-1 β and IL-18 in AGS cells. To assess effects of PFDA on the secretion of the cytokines that may be regulated by the inflammasome mechanism, we treated the gastric epithelial cell line AGS with PFDA and monitored IL-1 β and IL-18 productions by enzyme-linked immunosorbent assay (ELISA). As shown in Fig. 1a and b, cells incubated with PFDA had significantly promoted IL-1 β and IL-18 secretion in culture media compared with DMSO-treated control cells. PFDA enhanced IL-1 β production in culture media by more than 90% compared with control cells within 24 h; this ratio increased to 220% for IL-18. This increased yield of IL-1 β and IL-18 was verified by RT-qPCR of AGS cells (Fig. 1c and d). The results confirm that PFDA increased IL-1 β and IL-18 production in human gastric cells.

Moreover, the human hepatic cell line Bel7402, macrophage cell line THP-1 and normal murine hepatic cells were also used for testing IL-1 β and IL-18 production by ELISA after treatment with PFDA. As demonstrated in Fig. 1e,f,g,h,i and j, Bel7402, THP-1 and normal murine hepatic cells treated with PFDA raised IL-1 β and IL-18 production compared with DMSO-treated control cells. These results support the hypothesis that PFDA has an effect on IL-1 β and IL-18 production in human cells.

PFDA promotes caspase-1 activation in gastric epithelial cells via NLRP3 inflammasome. To determine if PFDA-promoted secretion of IL-1 β and IL-18 was a consequence of increased inflammasome activity, the level of active caspase-1 was examined in AGS concomitant with PFDA treatment. As shown in Fig. 2a, PFDA stimulated production of activate caspase-1.

To find out which inflammasome(s) were involved in PFDA's action, we performed real-time PCR to analyze the changes of mRNA levels of NLRP1, NLRP3 and NLRC4. The mRNA level of NLRP3 was increased by PFDA treatment and in a time-dependent manner (Fig. 2b). On the contrary, NLRP1 had no significant change in mRNA level in response to PFDA treatments (Fig. 2c), whereas NLRC4 could not be detected (no visible peak). This observation was consistent with an increase of NLRP3 protein level upon PFDA exposure as detected by Western blot (Fig. 2d). The dependence of cytokine secretion on NLRP3 was further verified by NLRP3 knockout mice, the promotions in IL-1 β and IL-18 production were abrogated in their stomach (Fig. 2e and f).

cIAP1 and -2, which are required for efficient caspase-1 activation by the inflammasomes, were involved in PFDA induced NLRP3 inflammasome activation. cIAP1 and cIAP2 are critical effectors of the inflammasomes and are required for inflammasome assembly and Caspase-1 activation. siRNA-mediated depletion of endogenous cIAP1 or cIAP2, alone or in combination, dramatically blunted caspase-1 activity¹⁷. Therefore, we thought to determine if cIAP1 and cIAP2 were involved in PFDA-promoted inflammasome activation. As shown in Fig. 3a and b, cIAP1 and cIAP2 mRNA levels were increased by PFDA, as shown by western blot at protein level (Fig. 3c and d). To confirm the role of cIAPs in PFDA-induced inflammasome activation, we investigated the effect of inhibition of cIAP2, by cIAP2 siRNA, on activation of caspase-1 and secretion of IL-1 β and IL-18. As shown in Fig. 3e,f and g, inhibition of cIAP2 abrogated PFDA-induced cIAP2 production and IL-1 β /IL-18 secretion.

NF κ B activity was enhanced in PFDA-treated gastric epithelial cells. NF κ B plays an important role in activating the priming step of inflammasomes and NF κ B inhibition results in a significant reduction of NLRP3 expression. To assess effects of PFDA on NF κ B activation, we treated AGS gastric epithelial cells with PFDA and monitored NF κ B activity with NF κ B reporter plasmid and Duo-Luciferase Assay Kit. As shown in Fig. 4a, cells incubated with PFDA had significantly increased NF κ B activity by 240% after PFDA treatment compared with DMSO-treated control cells. The enhanced NF κ B activity was verified by enhanced expression of c-Rel and p52 (Fig. 4b).

Promotion in secretion of IL-1 β and IL-18 and inflammation were detected in the stomach of PFDA-treated mice. To investigate the effects of PFDA on inflammasome activation and inflammation *in vivo*, we treated wild type C57BL/6 mice with PFDA in drinking water for 2 weeks and tested mice stomachs with ELISA. As shown in Fig. 5a and b, IL-1 β and IL-18 were increased in the stomach tissues of PFDA treated mice. Moreover, hematoxylin and eosin (HE) staining of stomach tissues of PFDA- or DMSO- treated C57BL/6 mice, showed disorganized alignment of cells and increased inflammatory cells infiltration (Fig. 5c and d). Therefore, these results demonstrate for the first time that PFDA regulates inflammasome assembly and inflammation in human cells and mice tissues.

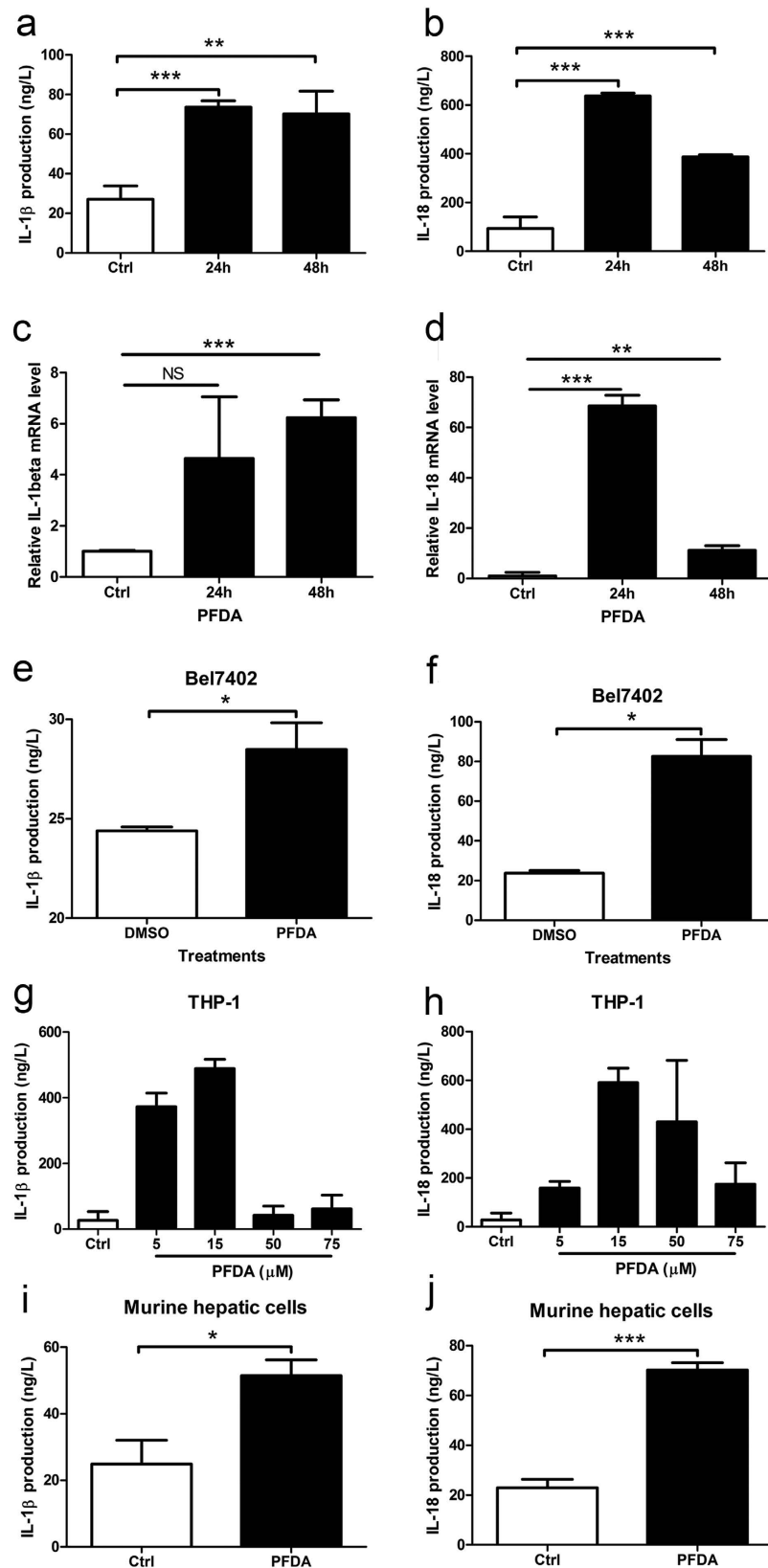


Figure 1. PFDA significantly stimulated IL-1 β and IL-18 production. IL-1 β and IL-18 secretions in culture media of AGS were increased after 5 μ M PFDA treatments (a and b), ANOVA = 0.011 and <0.0001 respectively); this increased secretion was verified by RT-qPCR (c and d); IL-1 β and IL-18 promotion was also detected in culture media of BEL7402 (e and f), THP-1 (g and h), ANOVA = 0.0005 and 0.01058 respectively) and normal murine hepatic cells (i and j). ELISA and RT-qPCR were performed as described in Materials and Methods.

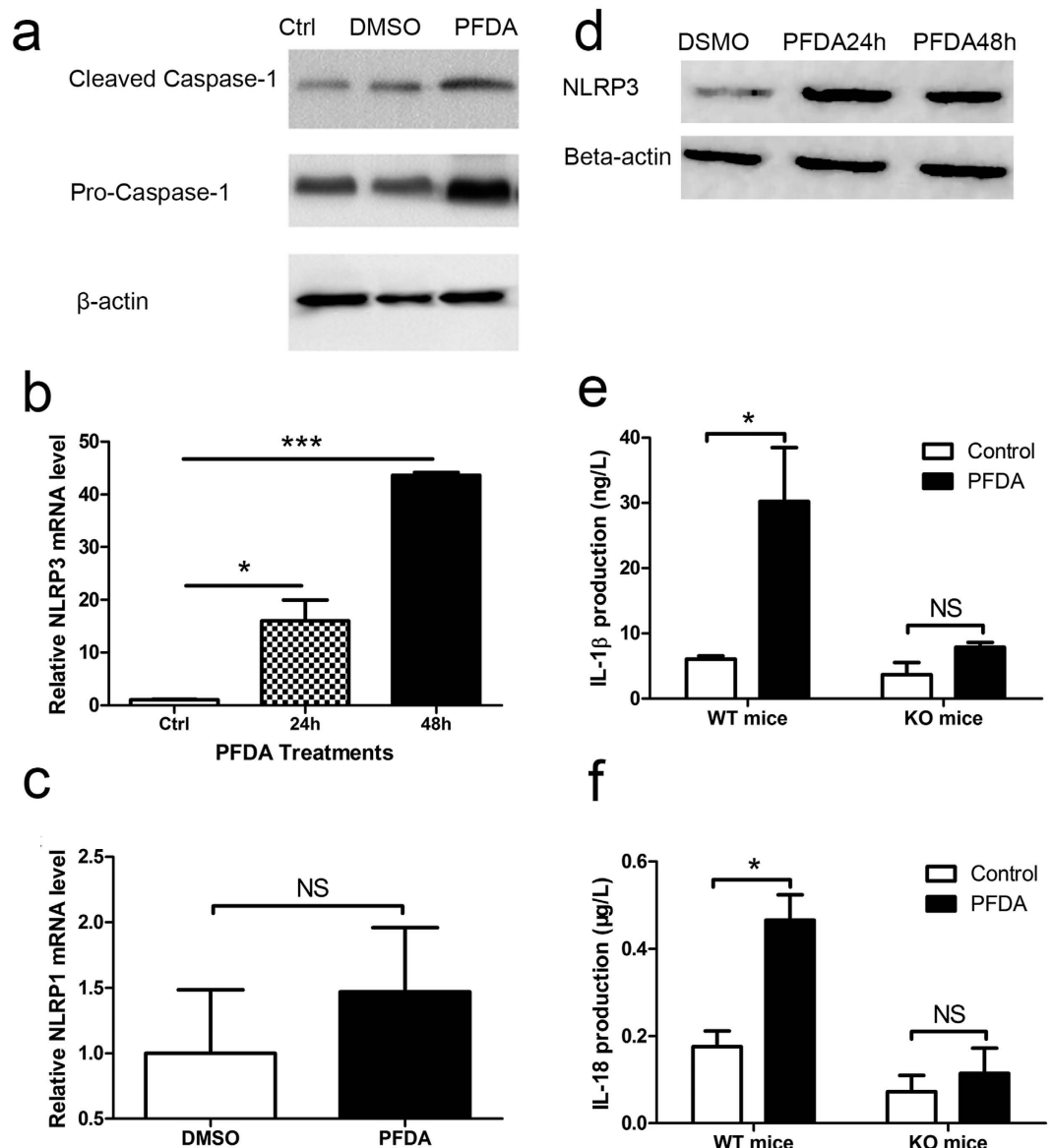


Figure 2. PFDA significantly enhanced NLRP3 inflammasome assembly. (a) Caspase-1 expression was increased after 5 μ M PFDA treatment for 24 h or 48 h in comparison with DMSO control; (b) the mRNA level of NLRP3 was increased by PFDA treatment and in a time-dependent manner (ANOVA <0.0001); On the contrary, NLRP1 had no significant difference in mRNA level in response to PFDA treatments (c); This mRNA increase of NLRP3 level upon PFDA exposure was verified by Western blot (d). The promotions in IL-1 β and IL-18 production were abrogated in their stomach of KO mice (e and f). RT-qPCR and Western blot were performed as described in Materials and Methods.

Discussion

Perfluorinated carboxylic acids or perfluorinated fatty acids (PFCAs) have been used for decades to make products that resist heat, oil, and water. Because they are used in the manufacture of nonstick cookware, fire-fighting foam, and many other industrial products^{18,19}, perfluorinated compounds can be detected globally in the environment^{6,20}, wildlife^{21–24} and humans^{25–30}. Continued monitoring of PFCAs is necessary since these chemicals persist in the environment and accumulate at much higher concentrations in human blood and organs because of their nondegradable nature and extremely long biological half-lives^{31,32}.

The reported cellular and physiological effects of PFCAs are associated with almost all systems of the human body; reports on the immunotoxicity of PFCs in humans are limited and controversial. On one hand, some reports suggested that exposure to PFCAs might be associated with immunosuppressive effects. Brieger *et al.* reported that Perfluorooctanoic Acid (PFOA) and Perfluorooctane Sulfonate (PFOS) were associated with reduced Natural killer (NK) cell activity and reduced the release of the pro-inflammatory cytokine Tumor necrosis factor (TNF)- α following lipopolysaccharide (LPS)-stimulation³³. Two recently published studies indicate that PFCAs may lower vaccine protection in children^{34,35}. Dong *et al.* and Zheng *et al.* discovered that the cytokine

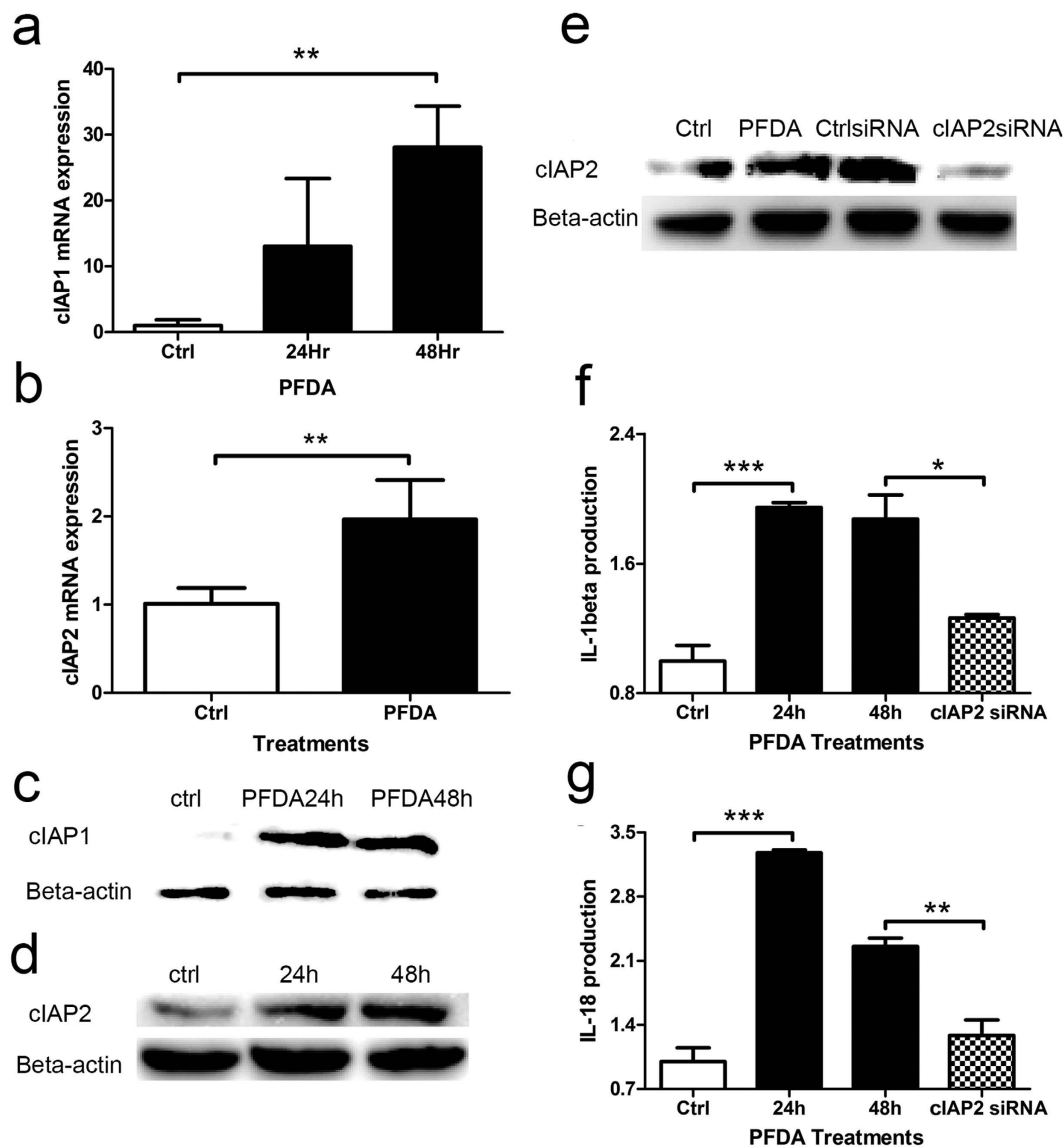


Figure 3. cIAP1 and -2 were involved in PFDA-associated NLRP3 inflammasome assembly promotion. (a and b), cIAP1 and -2 were activated by PFDA at mRNA level, this was evidenced by western blot at protein level (Fig. 3c and d). (e), inhibition of cIAP2 by siRNA abrogated PFDA-induced cIAP2 production; (f and g), inhibition of cIAP2 reduced IL-1 β and IL-18 secretion. ELISA, Western blot and RT-qPCR were performed as described in Materials and Methods.

balance favored helper T cell (Th)2 responses in experimental C57BL/6 male mice with short-term or subchronic PFOS exposure^{36,37}. On the other hand, some studies demonstrated that PFCAs act as an immunity enhancer. Wang *et al.* reported that prenatal PFOA and PFOS exposures positively correlated with cord blood IgE levels in boys³⁸. In agreement, Emmett *et al.* reported a slight increase in absolute monocyte counts of residents who lived in a water district contaminated with PFOA³⁹. Moreover, Ryu *et al.* found that utero-through-adulthood exposure of allergen-naïve mice to PFOA alone induced lung inflammation characterized by macrophage accumulation and IFN-expression, as well as Arylhydrocarbon Receptor (AHR), a hallmark symptom of asthma⁴⁰.

Despite recent advances in understanding the molecular mechanisms of perfluorinated environmental pollutants, many unanswered questions remain. PFDA is a straight chain ten-carbon PFCA that is structurally similar to fatty acids. *In vivo*, PFDA has highly potent and persistent toxicity⁵; it is several times as toxic as PFOA, so PFDA may be important for understanding the molecular mechanisms of toxicity. It is thus hoped that PFDA can assist in dissecting the sequence of events that ultimately results in tumor formation⁵. However, despite the evidence of PFDA toxicity, so far little is known of its effect on malignancy promotion and how it acts in inflammation. In this study, we evaluated the effects of PFDA on activation of the inflammasomes and inflammation regulation in the gastric cell line AGS. When added to cell cultures, PFDA significantly increased NLRP3 inflammasome activation compared with control cells. Moreover, the promotion in synthesis of IL-1 β and IL-18 were detected in the stomach of PFDA-treated mice. This report describes for the first time that PFDA regulates inflammasome assembly in gastric cells.

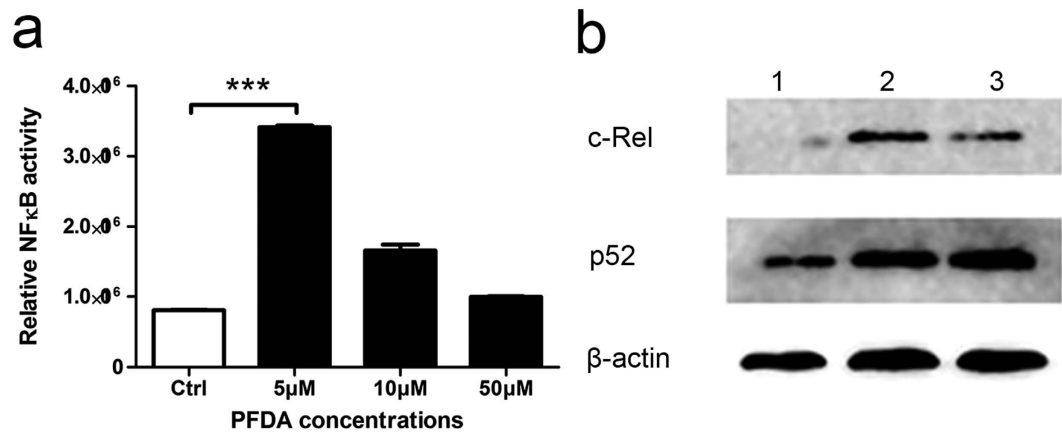


Figure 4. PFDA significantly enhanced NF κ B activity. (a) NF κ B activity was increased after PFDA treatment for 24 h in comparison with DMSO control and in a dose-dependent manner in response to PFDA, ANOVA <math><0.0001</math>; (b) This activation of NF κ B upon PFDA exposure was verified by the expression of c-Rel and p52: 1, DMSO control; 2, AGS with PFDA treatment for 24 h; 3, AGS with PFDA treatment for 48 h. NF κ B reporter assay was performed as described in the Kit instruction manual and Western blot were performed as described in Materials and Methods.

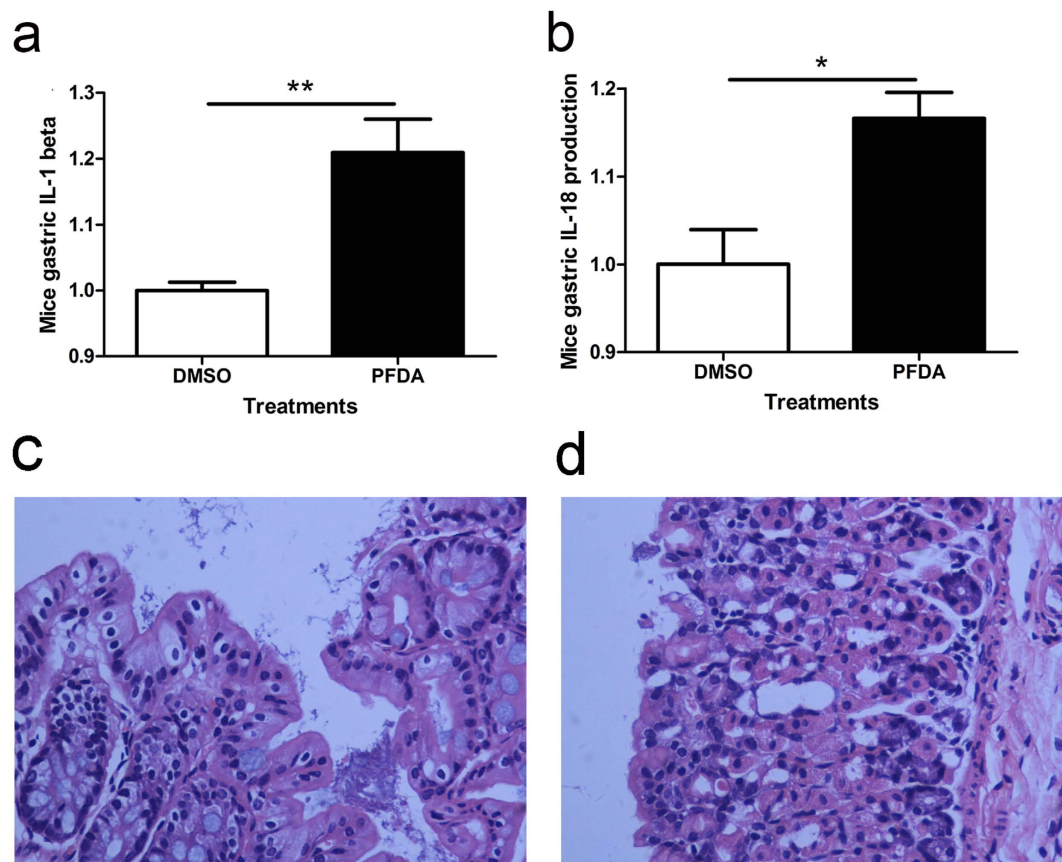


Figure 5. Promotion in secretion of IL-1 β and IL-18 and inflammation were detected in the stomach of PFDA-treated mice. IL-1 β and IL-18 secretions in mice gastric tissues were increased after PFDA treatments for 18 d at 25 mg/kg/d (a and b); Hematoxylin and Eosin (HE) staining of the stomach tissues of PFDA treated C57BL/6 mice showed disorganized alignment of cells and increased inflammatory cells infiltration (c and d). ELISA and HE staining were performed as described in Materials and Methods.

Proteins of the inhibitor of apoptosis protein (IAP) gene family have emerged as among the most important intrinsic inhibitors of apoptosis and could exert important roles in inflammation and innate immunity. They are characterized by the presence of a baculovirus IAP repeat (BIR) domain in one to three copies. cIAP1 and -2

function as ubiquitin ligases^{41,42} and were recently identified as important effectors of innate immunity, mediating critical steps in Nucleotide-binding oligomerization domain-containing protein (NOD) signaling⁴³. Moreover, cIAP1 and -2 were shown to mediate Mitogen-activated protein kinases (MAPK) activation downstream of Toll-like receptor 4 (TLR4) signaling by catalyzing degradative K48-linked ubiquitination of TNF Receptor Associated Factor 3 (TRAF3)⁴⁴. cIAP2-deficient mice have reduced cytokine amounts in response to LPS and are resistant to LPS endotoxemia⁴⁵. Labbe *et al.* reported that the cIAPs were critical effectors of the inflammasomes and were required for efficient caspase-1 activation. cIAP1, cIAP2, and the adaptor protein TRAF2 interacted with caspase-1-containing complexes and mediated the activating nondegradative K63-linked polyubiquitination of caspase-1¹⁷. Dagenais *et al.* reported that cIAP2-deficient mice had impaired activation of the regenerative inflammasome–interleukin-18 (IL-18) pathway⁴⁶. Moreover, the results of Mayer *et al.* revealed IAP antagonism as a profound anti-inflammatory principle *in vivo* and highlighted IAPs as important regulators of inflammatory processes in endothelial cells⁴⁷. In this study, we discovered that PFDA activated NLRP3 inflammasome through cIAP1/2, as a result of knockdown with cIAP2 siRNA, providing additional evidence that cIAP1/2 was involved in PFDA-induced inflammasome assembly.

NF κ B transcription factors play an important role in many physiological processes and diseases^{48,49}. IKK β preferentially phosphorylates the I κ B proteins, resulting in the release and activation of canonical NF κ B p50/p65 (RelA) and c-Rel/p65 complexes. IKK α homodimer-mediated phosphorylation of cytoplasmic NF κ B2/p100 results in partial proteasomal processing of p100 to produce mature p52, thereby activating the alternative pathway⁵⁰. NF κ B is thought to serve as the first signal that primes NLR and pro-IL-1 β expression^{51–53}. Qiao *et al.* demonstrated that NLRP3 promoter contains a putative NF κ B binding site and NF κ B inhibition resulted in a significant reduction of NLRP3 expression⁵⁴. Meanwhile, mounting evidence suggests that NLRP3 inflammasome formation is positively associated with NF κ B activity^{55,56}. All these findings implied that the pro-tumorigenic ability of NF κ B might be attributed to inflammasome activation.

cIAP1 and -2 are required for NF κ B signaling. Bertrand *et al.* revealed that cIAP1 and -2 were responsible for K63-linked polyubiquitination of receptor interacting protein 1 (RIP1), promoting NF κ B activation signaling⁵⁷. cIAP1 and cIAP2 also direct the ubiquitination of NF κ B-inducing kinase (NIK) to maintain the noncanonical NF κ B pathway^{58,59}. In this study, we discovered that PFDA promoted NF κ B reporter activity as well as enhanced c-Rel and p52 expression; thereby confirming activation of NF κ B signaling. Conclusively, this report describes for the first time that PFDA regulates inflammasome activation and inflammation in gastric cells.

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Author Contributions

S.L. wrote the main manuscript text; T.D. prepared Figure 5c and d and Figures related to K.O. mice; Z.F. prepared Figure 1e and f; Y.P. prepared Figure 3d and e; B.F. prepared Figure 3a, b and c; X.Z. and N.S. prepared the rest

Figures; R.Z. and X.W. provided with K.O. mice; M.H. and J.J. provided with some experimental reagents and laboratory facilities. All authors have reviewed the final version of the manuscript and approved this submission.

Additional Information

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